

# Mathematical modeling of temperature effect on algal growth for biodiesel application

S.M. Zakir Hossain<sup>1,\*</sup>, Nader Al-Bastaki<sup>2</sup>, Abdulla Mohamed A. Alnoaimi<sup>1</sup>, Husny Ezuber<sup>1</sup>, Shaikh A. Razzak<sup>3</sup>, and Mohammad M. Hossain<sup>3</sup>

<sup>1</sup> Department of Chemical Engineering, University of Bahrain, Zallaq, Kingdom of Bahrain

<sup>2</sup> Academic Affairs and Scientific Research, Kingdom University, Riffa, Kingdom of Bahrain

<sup>3</sup> Department of Chemical Engineering, King Fahd University of Petroleum and Minerals, Dhahran 31261, Saudi Arabia

Received: 7 November 2018 / Accepted: 13 March 2019

**Abstract.** Microalgae biomass is promising feedstock for the industrial production of biodiesel. Hence, research and development are required in various domains especially optimizations of growth conditions including temperature effect for mass scale operation (production of biomass, harvesting, extraction of lipid, etc). Since in middle east region, seasonal temperature variation and more rapid daily fluctuations are amenable to alter the growth kinetics of microalgae in outdoor culture and hence affect algae biomass production efficiency. Therefore, in this study, a mathematical model was developed to calculate how the algae sp. (*Chlorella kessleri*) will react at different temperatures. The model integrates Monod model and Arrhenius equation, and as such it describes the relationship of algal growth rate with culturing temperature and limiting nutrient concentration. The apparent activation energy and pre-exponential factors were calculated to be 2537 cal/mol and 0.0077 day<sup>-1</sup>, respectively. The developed models could be useful to anticipate the effective impacts of temperature on outdoor algae culture.

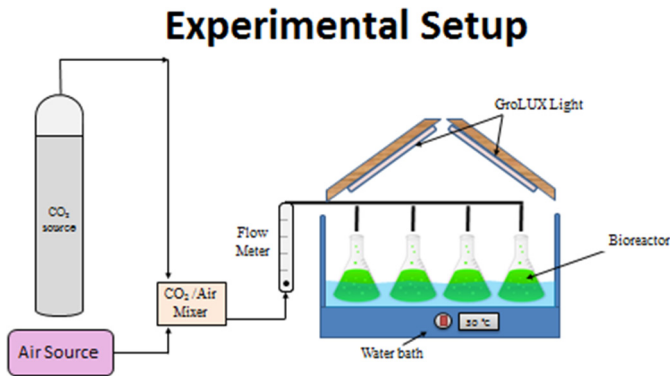
## 1 Introduction

The trend in recent years is replacing fossil fuels by liquid biofuels. First generation biofuels have achieved the required production rate economically. The energy source for this generation was food and oil crops including oil, sugar, maize and animal fats. It is obvious that the generation and consumption of liquid biofuels will remain the same and might grow in the near future. In general, the utilization of first-generation biofuels has formed several arguments, mainly because of their effect on international food markets and on food security, particularly with respects to the most vulnerable regions of the global economy. Relevant questions were appeared about their effectiveness and sustainability of their production. Second-generation biofuels produced fuels from the whole plant matter of dedicated energy crops or agricultural deposits, jungle gathering remains or lumber processing waste, rather than from food crops. However, the production technologies for the second-generation biofuel in the most part have not met the scales for commercial use and not attract with the market demand which has so far prevented any substantial exploitation [1]. For a

technically and economically feasible fuel supply, some conditions must be satisfied: relatively cheap, should need low or no additional land usage, environmentally friendly and no contribution to air pollution, and should use minimal amount of water. Microalgae exploitation could achieve these requirements and therefore makes an important influence to meeting the main energy demand and, in the mean, while providing environmental assistances [2]. Microalgae are all unicellular and simple multicellular microbes, counting both prokaryotic microalgae, i.e. cyanobacteria (*Chloroxybacteria*), and eukaryotic microalgae, e.g. green algae (*Chlorophyta*), red algae (*Rhodophyta*) and diatoms (*Bacillariophyta*) [2–4]. Algae culture has several potential advantages that include: (i) uses non-arable land for their culturing so does not vulnerable to food security, (ii) provides high oil yield per acre (using microalgae yield of 10 000-gallon oil per acre compared with 625-gallon oil per acre for oil palm), (iii) has fast exponential growth and (iv) uses CO<sub>2</sub>, nitrogen, phosphorus and brackish/waste water during growth. As such, algae culture can contribute to CO<sub>2</sub> biofixation, wastewater treatment and source of biofuel [5–12].

Generally, temperature, nutrient (nitrogen and phosphorus) concentration, light intensity, initial CO<sub>2</sub> concentration and initial inoculum size are the parameters that affect the microalgae growth rate. Among these,

\* e-mail: [zhossain@uob.edu.bh](mailto:zhossain@uob.edu.bh)



**Fig. 1.** Schematic diagram of the experimental setup used to cultivate microalgae.

temperature and nitrogen concentration are considered the main parameters. It is well known that models of cell culture are useful tools to predict growth kinetics as well as better information in aquatic resource management [13–15]. To our knowledge there is no mathematical model for *Chlorella kessleri* (one of the green microalgae sp.) available in literature. Therefore, this paper develops a mathematical model of *C. kessleri* growth at different temperature and nitrogen concentration. For this, both Monod model and Arrhenius equation were applied. The development of this model could open the opportunity to propose models of automated bioprocess control and optimize productivity at large scale, which in turn helpful for biodiesel production.

## 2 Materials and methods

### 2.1 Chemical and reagents

All the solvents and chemicals used were obtained from Sigma–Aldrich, unless stated otherwise.

### 2.2 Algae species and growth conditions

The microalgae species (*C. kessleri*) was obtained from Algae depot in the USA. The algae species was cultured in a media called Bold's Basal Medium (BBM), which was prepared based on the recipe shown in the Supplementary Table S1. The pH of the medium was adjusted to 7 by NaOH (0.1 M). The experimental setup for this study is shown in Figure 1. To avoid the lag phase, the microalgae species was initially pre-cultured and then moved into the photobioreactors (PYREX 1L Erlenmeyer flasks, working volume was 500 mL each). The initial concentrations of the microalgae strain were  $2.2 \times 10^7$  cells mL<sup>-1</sup>. The photobioreactors were arranged on a bench, where GroLUX fluorescents light bulbs (light intensity of  $65 \mu\text{mol m}^{-2} \text{s}^{-1}$  measured by an LI-250 Light Meter) were placed above the water bath that regulates the culturing temperature. CO<sub>2</sub>–air mixture (96% air and 4% CO<sub>2</sub>) was fed at the bottom of the batch reactors.

### 2.3 Measurement of algae biomass

For growth analysis, the algae samples (1 mL each) were obtained in every 24 h intervals. The cell concentration was calculated by measuring the absorbance at 690 nm using a UV-spectrophotometer (UNICAM). The dry biomass values (g/L) were obtained after vacuum filtration of 10 mL sample at every 3 days, with drying at 55 °C for 24 h. The biomass concentration (g/L) was estimated based on a biomass-optical densities standard curve (Supplementary Fig. S1).

### 2.4 Measurement of algae growth kinetic parameters

The specific growth rate ( $\mu$ , or SGR, day<sup>-1</sup>) and biomass productivity ( $P$ , g<sub>dw</sub> L<sup>-1</sup> day<sup>-1</sup>) were determined from cell concentration and dry biomass weight data by using the following equations (1) and (2), respectively [15–20]:

$$\text{Specific growth rate, } \mu = \frac{\ln\left(\frac{N_2}{N_1}\right)}{t_2 - t_1} \quad (1)$$

$$\text{Biomass productivity, } P = \frac{X_t - X_0}{t_x - t_0} \quad (2)$$

where  $N_1$  and  $N_2$  are the biomass concentrations (g/L) at the beginning  $t_1$  and at the end  $t_2$  of the exponential growth phase, respectively;  $X_t$  is the biomass concentration (g/L) at the end of the cultivation period  $t_x$  and  $X_0$  the initial biomass concentration (g/L) at  $t_0$  (day).

### 2.5 Determination of nitrogen removal efficiency

The removal of nitrogen efficiency was estimated by quantifying the level of removal (obtained by subtracting from initial to final values) from the culturing medium. The filtrate (obtained while performing biomass dry weight) was subjected to nitrate test using Chromo tropic acid method with acid persulfate digestion respectively according to procedure given in Hach Company, 2015 [21].

### 2.6 Development of mathematical models and validation

Mathematical model was developed to determine the quantitative linkage between environmental parameters (temperature, nitrate concentration in the medium) and cell growth kinetics. For simplification, prior to formulate model, the following assumptions were made: (i) light intensity was kept constant, (ii) initial inoculum size was kept constant and (iii) death kinetics were ignored. First, the effect of limiting nutrient (nitrate) on the growth of microalgae was described by the Monod model as shown in equation (3) [22].

$$\mu = f(S) = \mu_m \left[ \frac{S}{K_s + S} \right] \quad (3)$$

where  $\mu$  is the specific growth rate,  $\text{day}^{-1}$ ;  $\mu_m$  the maximum specific growth rate,  $\text{day}^{-1}$ ;  $S$  the limiting nutrient concentration,  $\text{mg/L}$  and  $K_s$  the half saturation coefficient,  $\text{mg/L}$ .

The maximum specific growth rate ( $\mu_m$ ), which appears in the Monod model, is not a function of limiting nutrient concentration, rather, it is a function of temperature and light intensity. Since the light intensity was assumed constant, the maximum specific growth rate can be a function of temperature only and it may be described fairly by the Arrhenius equation (4) [22].

$$\mu_m = Ae^{-E/RT} \quad (4)$$

where  $A$  is a constant,  $\text{day}^{-1}$ ;  $E$  the activation energy,  $\text{cal/mol}$ ;  $R$  the universal gas constant,  $\text{cal/K/mol}$  and  $T$  the temperature,  $\text{K}$ .

Upon combination of equations (3) and (4), equation (5) was formulated, which can describe the growth of microalgae as a function of temperature and limiting nutrient concentration.

$$\mu = Ae^{-E/RT} \left[ \frac{S}{K_s(T) + S} \right]. \quad (5)$$

The half-saturation coefficient is an empirical value which can be found easily by plotting specific growth rate ( $\mu$ ) vs. nutrient concentration ( $S$ ). It is important to note that  $K_s$  is the substrate concentration at half maximal specific growth rate ( $\mu_m$ ) and it depends on environmental conditions such as the temperature.

For model validation, theoretical specific growth rates (computed using the model equation) vs. nutrient concentrations ( $S$ ) were plotted for different temperature. These figures were compared with those obtained in experiment.

### 3 Results and discussion

#### 3.1 Growth profile of microalgae sp. *Chlorella kessleri*

To characterize the time-resolved growth phase, it is important to explore the growth behavior of microalgae sp. *C. kessleri* in BBM at different culturing temperature. The population of *C. kessleri* grown at  $25^\circ\text{C}$  in a batch photobioreactor containing BBM is shown in Figure 2. One can see from this figure that for 3 h incubation, the growth of *C. kessleri* was slow. The growth then continued to increase until day 9 before touching the stationary phase. The logarithmic growth phase was observed within 3–9 days and thereby maximum specific growth rates at different culturing temperature were determined in this region. Limiting nutrients concentration in the form of nitrate ( $\text{NO}_3^-$ ) was monitored by plotting the nitrate concentration ( $\text{mg/L}$ ) vs. time (day). The decreasing pattern of consumption of limiting nutrient, nitrate by *C. kessleri*, is shown in Figure 3. It is important to note that different growth and nitrate consumption patterns were observed at different culturing temperatures. As such, it is important to develop a model that describes the temperature effect on microalgal growth.

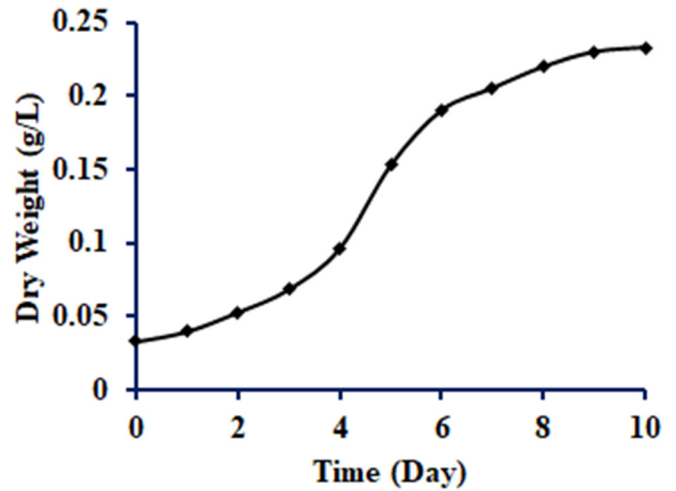


Fig. 2. Growth curve for microalgae sp. *Chlorella kessleri* in BBM at  $25^\circ\text{C}$  in a batch bioreactor.

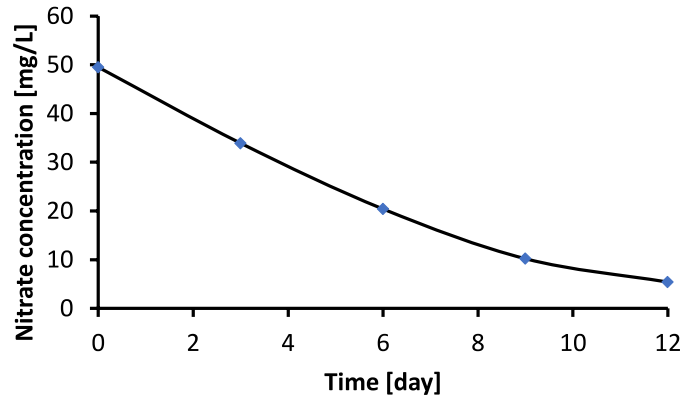


Fig. 3. Consumption of nitrate with time by *Chlorella kessleri* in BBM at  $25^\circ\text{C}$ .

#### 3.2 Development of mathematical model for algal growth

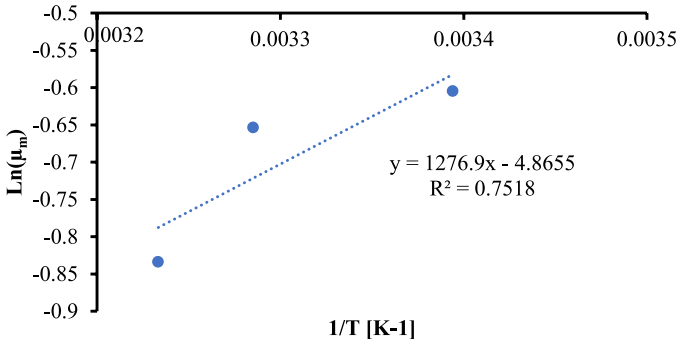
In order to obtain the parameters of pre-exponential factor ( $A$ ) and activation energy ( $E$ ) in the Arrhenius model, maximum specific growth rate ( $\mu_m$ ) was calculated for each temperature and the data were shown in Table 1. Figure 4 shows a plot of  $\ln(\mu_m)$  vs.  $1/T$ . It was a straight line with the  $R^2$  value of 0.7621, indicating good fit. Since in biological system, the  $R^2$  value should be more than 0.75 to validate the empirical model [23]. The  $A$  and  $E$  were calculated to be  $0.0077 \text{ day}^{-1}$  and  $2537 \text{ cal/mol}$ . Thus, Arrhenius model was rewritten with these values as:

$$\mu_m = 0.0077 \exp\left(\frac{1277}{T[\text{K}]}\right). \quad (6)$$

It is essential to note that negative activation energy was observed. The probable reason behind this is that increasing the temperature causes increase in the growth of algae to a certain level then decrease in the growth. The

**Table 1.** Data used to fit the Arrhenius model.

$T$ (°C)	$\mu_m$ (day <sup>-1</sup> )	$\ln(\mu_m)$	$1/T$ (K <sup>-1</sup> )
25	0.55	-0.605	0.0034
35	0.52	-0.653	0.0033
40	0.44	-0.834	0.0032

**Fig. 4.** A plot of  $\ln(\mu_m)$  vs.  $1/T$ .**Table 2.** A comparison between experimental and predicted maximum specific growth rates.

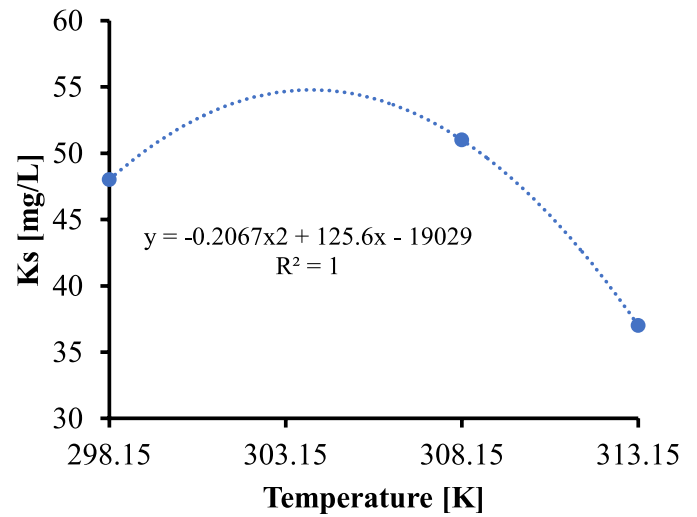
$T$ (°C)	$\mu_{\text{exp}}$ (day <sup>-1</sup> )	$\mu_{\text{Arr}}$ (day <sup>-1</sup> )	%Error <sub>exp/Arr</sub>
25	0.55	0.56	2.2%
35	0.52	0.49	6.6%
40	0.44	0.45	4.7%

model (Eq. (6)) was used to anticipate the maximum specific growth rates. A comparison between experimental and predicted values of the maximum specific growth rates in terms of % errors is shown in Table 2. The maximum error between predicted and experimental specific growth rate was found less than 7%. Thus, the developed model was reliable.

The next step was to incorporate both temperature and nutrient concentration into one model. The nutrient limitation was modeled by finding the empirical coefficient of the Monod equation, which is the half-saturation coefficient ( $K_s$ ). The half saturation coefficient was found graphically from a plot of specific growth rate ( $\mu$ ) vs. nutrient concentration ( $S$ ). Since the half saturation coefficient ( $K_s$ ) is a function of temperature, its values were determined for each temperature as shown in Table 3. A relationship between the half saturation coefficient and temperature is obtained by plotting  $K_s$  (mg/L) vs. temperature (K) as depicted in Figure 5. Apparently, a second order polynomial relationship (as seen in Eq. (7)) was obtained for describing the effect of temperature on the half saturation coefficient. It is obvious from the figure that at moderate temperature the value of  $K_s$  was relatively

**Table 3.** Half saturation coefficient at different temperatures.

$T$ (°C)	$K_s$ (mg/L)
25	48
35	51
40	37

**Fig. 5.** The effect of temperature on the half saturation coefficient ( $K_s$ ).

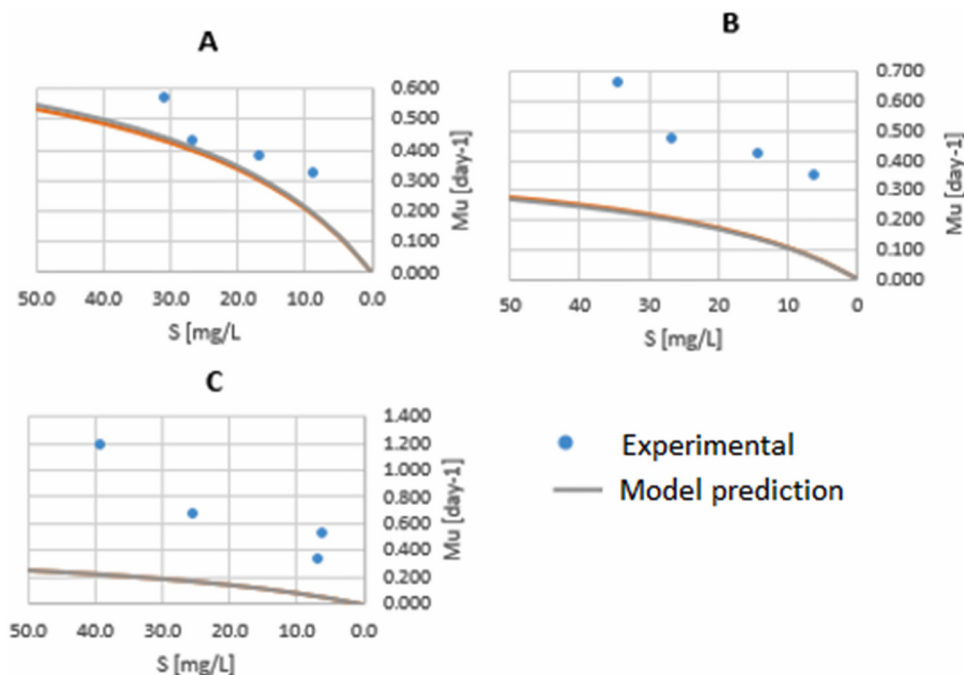
higher than that of both low and high temperature. Then, equation (7) was incorporated into the Monod model (Eq. (3)), and a resultant model was obtained (Eq. (8)), that describes the specific growth rate as a function of limiting nutrient concentration.

$$K_s = K_s(T) = -0.2067T^2 + 125.6T - 19029 \quad (7)$$

$$\mu = \mu_m \left[ \frac{S}{(-0.2067T^2 + 125.6T - 19029) + S} \right]. \quad (8)$$

Finally, an integrated model was made (as shown in Eq. (9)) by combining equations (8) and (6) and it describes the algal growth as a function of temperature and limiting nutrient concentration. This model was used to predict the specific growth rate for any temperature and nutrient concentration. The optimum temperature and nutrient concentration that will maximize the specific growth rate can also be measured.

$$\mu = 0.00771 \exp\left(\frac{1276.9}{T[K]}\right) \times \left[ \frac{S}{(-0.2067T^2 + 125.6T - 19029) + S} \right]. \quad (9)$$



**Fig. 6.** A comparison between experimental and model prediction of specific growth rate ( $\mu$ ) at different temperatures: (A) 25 °C, (B) 35 °C and (C) 40 °C.

### 3.3 Model validation

The maximum specific growth rate was determined by model equation (6) that agrees satisfactorily with the experimental data for all temperature. For integrated model (Eq. (9)) validation, predicted specific growth rates vs. nutrient concentrations ( $S$ ) were plotted for different temperatures, separately as shown in Figure 6A–C. These figures were compared with those obtained in experiment in the same figures. The predicted data are relatively deviated from the experimental values indicating fairly predictions in specific growth rate because these models are quadratic equations and are possibly inadequate to define the algal growth effectively in all kinetic regimes. Besides this model is solely a function of temperature and limiting nutrient concentration. Other important factors (e.g., light intensity, initial inoculum size, decay rate) that could possibly affect the specific growth rate of microalgae were ignored during model development.

## 4 Conclusions

In the present study, a mathematical model was formulated by combining Monod model and Arrhenius equation based on some assumptions. It describes the relationship of algal growth rate with culturing temperature and limiting nutrient concentration. This model was used to predict microalgal (*C. Kessleri*) specific growth rate at different temperatures. The apparent activation energy and pre-exponential factors were calculated to be 2537 cal/mol and 0.0077 day<sup>-1</sup>, respectively. The developed model agreed fairly with experimental data and could be useful to visualize the effective impacts of temperature on outdoor

algae culture. However, to get a perfect prediction of growth rate by the model, it is important to develop a comprehensive model considering all possible factors such as culture temperature, nutrient (nitrate and phosphate), light intensity, initial inoculum size, CO<sub>2</sub> uptake rate and death kinetics. Our research is underway in this direction.

### Supplementary material

**Supplementary Table and figure.** The Supplementary Material is available at <https://www.rees-journal.org/10.1051/rees/2019005/olm>.

The authors would like to gratefully acknowledge the support provided by King Abdulaziz City for Science and Technology (KACST) through the Science & Technology Unit at King Fahd University of Petroleum & Minerals (KFUPM) for funding this work through project No. NSTIP # 13-WAT096-04 as part of the National Science, Technology and Innovation Plan.

## References

1. B. Liam, P. Owende, Renew. Sustain. Energy Rev. **14**, 557 (2010)
2. B. Wang, Y. Li, N. Wu, C.Q. Lan, Appl. Microbiol. Biotechnol. **79**, 707 (2008)
3. J. Cheng, Y. Huang, J. Feng, J. Sun, J. Zhou, K. Cen, Bioresour. Technol. **144**, 321 (2013)
4. M.G. de Moraes, J.A.V. Costa, Biotechnol. Lett. **29**, 1349 (2007)
5. S.M.Z. Hossain, S.A. Razzak, M.M. Hossain, Chem. Eng. Technol. **41**, 1313 (2018)

6. S.M.Z. Hossain, A. Alnoaimi, S.A. Razzak, H. Ezuber, N.A. Bastaki, M. Safdar, S. Alkaabi, M.M. Hossain, *Can. J. Chem. Eng.* **96**, 1903 (2018)
7. M.A. Kazeem, S.M.Z. Hossain, S.A. Razzak, M.M. Hossain, *Chem. Prod. Process Model.* **13**, 20170082 (2018)
8. P.M. Schenk, S.R. Thomas-Hall, E. Stephens, U.C. Marx, J. H. Mussgnug, C. Posten, O.R.J. Kruse, B.D. Hankamer, *Bioenergy Res.* **1**, 20 (2008)
9. G.C. Dismukes, D. Carrieri, N. Bennette, G.M. Ananyev, M.C. Posewitz, *Curr. Opin. Biotechnol.* **19**, 235 (2008)
10. K.B. Cantrell, T. Ducey, K.S. Ro, P.G. Hunt, *Bioresour. Technol.* **99**, 7941 (2008)
11. Y. Chisti, *Biotechnol. Adv.* **25**, 294 (2007)
12. S.A. Razzak, M.M. Hossain, R.A. Lucky, A.S. Bassi, H. De Lasa, *Renew. Sustain. Energy Rev.* **27**, 622 (2013)
13. D. Zhang, P. Dechatiwongse, E.A. del Rio-Chanona, G.C. Maitland, K. Hellgardt, V.S. Vassiliadis, *Algal Res.* **9**, 263 (2015)
14. J.E. Bissinger, D.J.S. Montagnes, *Limnol. Oceanogr.* **53**, 487 (2008)
15. C. Ocampo-López, S. Colorado-Arias, M. Ramírez-Carmona, *J. Appl. Res. Technol.* **13**, 498 (2015)
16. S.F. Mohsenpour, B. Richards, N. Willoughby, *Bioresour. Technol.* **125**, 75 (2012)
17. P. Das, W. Lei, S.S. Aziz, J.P. Obbard, *Bioresour. Technol.* **102**, 3883 (2011)
18. M.G. De-Morais, J.A.V. Costa, *J. Biotechnol.* **129**, 439 (2008)
19. D. Tang, W. Han, P. Li, X. Miao, J. Zhong, *Bioresour. Technol.* **102**, 3071 (2011)
20. Y.S. Yun, S.B. Lee, J.M. Park, C.I. Lee, J.W. Yang, *J. Chem. Technol. Biotechnol.* **69**, 451 (1997)
21. Chromotropic Acid Method, Test 'N Tube™ Vials, Hach Company, ed. 10 (2015)
22. J.C. Goldman, E.J. Carpenter, *Limnol. Oceanogr.* **19**, 756 (1974)
23. P.D. Haaland, *Experimental Design in Biotechnology* (CRC Press, Taylor & Francis Group, Boca Raton, 1989)

**Cite this article as:** S.M. Zakir Hossain, Nader Al-Bastaki, Abdulla Mohamed A. Alnoaimi, Husny Ezuber, Shaikh A. Razzak, Mohammad M. Hossain, Mathematical modeling of temperature effect on algal growth for biodiesel application, *Renew. Energy Environ. Sustain.* **4**, 8 (2019)